RESEARCH ARTICLE

PHYTOCHEMICAL ANALYSIS AND ANTIBACTERIAL EFFICACY OF OCIMUM GRATISSIMUM EXTRACTS ON MULTI-DRUG RESISTANT BACTERIA

Adebesin, IO¹, Kolapo, KT², Amoka, SO³, Olubunmi, OH⁴, Olomoko, CR.⁵, Balogun, AO⁶, Ganiyu, LT⁷

Received: 30 Oct, 2023/Revision: 03 Nov, 2023 / Accepted: 30 Nov, 2023

Abstract: The continuous indiscriminate use of antibiotics in treating animals and human diseases has led to an increase in the ability of bacteria to resist the effects of various antibiotics in use, and this has led to the study and screening of plants for their ability to treat various diseases. In this study, the bioactive components and the antibacterial efficacy of Ocimum gratissimum ethanolic and methanolic leaf extracts against Staphylococcus aureus, Pseudomonas aeruginosa, Klebsiella pneumoniae, Escherichia coli S. marcescens and B. cereus were determined using the Analytical Methods Committee's standard method and agar well diffusion method respectively. The phytochemical components, antibacterial activity, MIC, and MBC of O. gratissimum were determined against these isolates. The qualitative phytochemical analysis of the extracts revealed the presence of saponins, alkaloids, phenols, tannins, steroids, flavonoids, anthraquinones, terpenoids, glycoside and phlobatannins in both or either of the extracts. The result revealed ethanolic extract to be more potent than methanolic extract by producing inhibition ranges of 15-35mm, 8-27mm, 7-26mm, 9-27mm, 5-24mmm, 13-32mm (ethanol) and 12-33mm, 11-31mm, 8-23mm, 12-25mm, 8-26mm, 6-26mm (methanol) against S. aureus, K. pneumoniae, P. aeruginosa, E. coli, S. marcescens and B. cereus at 12.5-200mg/ml respectively. The MIC and MBC ranged between 12.5 - 50mg/ml and 50-100mg/ml for both extracts respectively. This study revealed that O. gratissimum possesses antibacterial efficacy on the selected isolates even at low concentrations; thereby its use is recommended for the treatment of various diseases caused by these organisms.

KEYWORDS: antibacterial efficacy, MIC, MBC, phytochemicals, resistance.

Corresponding Author: Ibraheem O. Adebesin,

Environquest Integrated Environmental Solution, Lagos, Nigeria.

Email-ibraheemolakunle7@gmail.com



^{1,4}Environquest Integrated Environmental Solution, Lagos, Nigeria.

²International Institute of Tropical Agriculture (IITA), Ibadan, Nigeria.

³⁵⁶Department of Microbiology, Faculty of Life Sciences, University of Ilorin, Ilorin, Nigeria.

⁷Ladoke Akintola University of Technology, Ogbomoso, Nigeria.

INTRODUCTION:

The daily increase in resistance of bacteria to conventional antibiotics has posed a serious challenge to researchers, pharmaceutical companies, and health practitioners worldwide. These resistant bacteria are usually transferred to humans from animals through the ingestion of contaminated food, leading to the exchange of resistant genes between these pathogenic strains and body microbiota in the host. Antibiotic resistance in bacteria often results from target site modification, overproduction of the target sites, decrease in cell membrane permeability, natural selection, and efflux pump of the antibiotics. This resistance ensures their survival in the presence of these bactericidal and bacteriostatic substances in low, mid, and high concentrations; antibacterial. Hence, the need and inspiration to search for new antimicrobial agents from other sources like plants to overcome the problem of resistance and improve the quality of life. For years, new antibiotics have not been developed despite the large number of bacteria that have become resistant to almost all antibiotics in use for treating various bacterial infections in animal husbandry and humans. In Africa, herbs have been in use for the treatment of ailments such as cough, fever, diarrhea, and sore throat for ages.[1] It was reported that Vernonia amygdalina, Carica papaya and Carthamus tinctorius possess antimicrobial efficacy against multidrug resistant bacteria and fungi. [2-4] It is therefore worthy to study the antibacterial efficacy of Ocimum gratissimum on some multi-drug resistant bacteria that are becoming resistant to various antibiotics and posing threats to an individual's health. The increase in diseases caused by multi-drug resistant bacteria and the speed at which antibiotic-resistant genes are spreading in the environment, demand conservative efforts in searching for alternatives that are effective, available, affordable, and non-toxigenic to humans and animals.

Ocimum gratissimum is a perennial plant that is widely distributed in Africa and Asia. It belongs to the family Lamiaceae with 1-3m in height, dark brown stem,

narrow, small, and oval-shaped leaves. In Nigeria, it is called "scent leaf" due to the fascinating aroma it produces that encourages its use as a spice in food^[5] and when consumed, it serves as medicine for treating diseases in humans and animals.^[6] This plant has been reported to possess antibacterial and antidiarrheal properties, antifungal, antidiabetic, treat blocked nostrils, regulate menstruation, ear infections and cough and remedy for constipation.[7-11] The essential oil contained in the leaves and flowers of O. gratissimum has been used in the preparation of tea, perfume, and infusion by Nigerians.[12] The whole plant is also used by Indians to treat headache, inflammation and pyretic.[13] It is also used by Nigerians to treat epilepsy, high fever, and diarrhea. [14] This study is conducted to evaluate phytochemicals in O. gratissimum leaves and its antibacterial efficacy against some multi-drug resistant bacteria.

MATERIALS AND METHODS:

Collection of Plant Materials

Fresh leaves of the *O. gratissimum* were collected from the environment. Identification and authentication of the leaves were carried out at the herbarium section of the Department of Plant Biology, University of Ilorin, Ilorin, Nigeria.

Preparation of Crude Extracts

The leaves were dried in an open space and ground into powder form. Fifty grams (50g) of the ground leaf material was soaked in 250ml of ethanol and methanol for 48 hours separately. The mixtures were sieved to remove debris. The filtrates were then filtered through Whatman No. 1 filter paper and opened to air for the solvent to evaporate to get the crude extract [15]. The crude methanol and ethanol extracts were stored at 4°C until needed.

Preparation of Concentration of Plant Extract

Two grams (2g) each of ethanol and methanol extracts were added to the corresponding 10ml ethanol and

methanol separately to give an equivalent concentration of 200mg/ml while other lower concentrations (100, 50, 25 and 12.5mg/ml) were obtained by double fold dilution method of previous concentration.^[15]

Collection and Maintenance of the Test Organisms

Pure cultures of two Gram-positive bacterial isolates (Staphylococcus aureus and Bacillus cereus) and four Gram-negative bacterial isolates (Klebsiella pneumoniae, Pseudomonas aeruginosa, Escherichia coli and Serratia marcescens) were obtained from the Department of Microbiology, University of Ilorin and were re-identified by biochemical tests. The cultures were stocked on nutrient agar slants before they were sub-cultured onto freshly prepared nutrient agar plates for antimicrobial analysis.

Qualitative Phytochemical Analysis

The presence of bioactive agents such as saponins, alkaloids, phenols, tannins, steroids, flavonoids, anthraquinones, terpenoids, glycoside and phlobatannins in the extracts was determined according to the standard methods for the analysis of phytochemicals by Analytical Methods Committee of Royal Society of Chemistry.^[16]

Antibacterial Activity of *O. gratissimum* Extracts on Multi-drug Resistant Bacteria

The antibacterial activity of ethanol and methanol extracts was determined by the agar well diffusion method. A flamed inoculating loop was used to pick a distinct colony from 24-hour old culture on nutrient agar into 10ml of sterile 0.85% normal saline in the test tube with gentle shaking to make the culture homogenized. The saline turbidity is then compared with 0.5 McFarland turbidity until it matches to obtain 10⁷CFU/ml. 0.1ml of the standardized aliquot was inoculated onto solidified Mueller Hinton agar, spread evenly by swap stick, and allowed to dry. A 4mm corkborer was used to make two wells on the plate. 0.1ml of the extract was measured into these wells. The antibacterial activity of the extracts' concentrations

against each isolate was determined by measuring the zone of inhibition around the well with meter rule in millimeters after 24 hours of incubation at 37°C. [17]

Antibiotics Susceptibility Testing of the Test Organisms

A swab stick moistened with standardized inoculum was used to spread the surface of solidified Mueller Hinton agar in a way that confluence growth can be obtained after incubation. A sterile forceps was used to place antibiotic discs on the surface of the dry inoculated plate. The diameter of inhibition around each disc was measured in millimeters after incubation at 37°C for 24 hours and interpreted as resistant, intermediate, or susceptible.^[18]

Determination of Minimum Inhibitory Concentration (MIC)

Broth dilution method was used to determine the minimum inhibitory concentration (MIC) of ethanolic and methanolic extracts against each of the test isolates at varying concentrations of 200, 100, 50, 25 and 12.5mg/ml. 1ml of the extract was introduced into 10ml nutrient broth inoculated with 0.1ml of the standardized organism and the control tube was made the same way without the test organism. The tube with the lowest concentration of no growth or having the same turbidity as the control tube after incubation at 37°C for 24 hours was recorded as the MIC of that bacteria. [19]

Determination of Minimum Bactericidal Concentration (MBC)

MBC of the extracts was determined by sub-culturing from tubes having the same turbidity as the control tube from the MIC test. 0.1ml of the 24-hour culture was inoculated onto solidified nutrient agar plates spread by a sterile glass spreader and incubated at 37°C for 24 hours. The minimum concentration at which there was no bacterial growth on the plates was recorded as the MBC for the isolate. [19]

Statistical analysis

The results were expressed as mean \pm SE of duplicate samples. Statistical analysis was performed by one-way ANOVA of SPSS version 20. The obtained results were considered statistically significant at p \leq 0.05.

RESULTS:

Effects of Extraction Solvent on Qualitative Phytochemical Constituents of *O. gratissimum* Leaves

The qualitative analysis of *O. gratissimum* ethanolic leaf extract revealed the presence of saponins, alkaloids, phenols, tannins, steroids, flavonoids, terpenoids, glycoside and phlobatannins while the methanolic extract revealed the presence of saponins, alkaloids, phenols, tannins, steroids, flavonoids, anthraquinones and glycoside (**Table 1**).

Table 1: Bioactive components of O. gratissimum.

Phytochemicals	Ethanol extract	Methanol extract
Alkaloids	+	+
Phenols	+	+
Saponins	+	+
Steroids	+	+
Flavonoids	+	+
Terpenoids	+	-
Glycoside	+	+
Phlobatannins	+	-
Tannins	+	+
Anthraquinones	-	+

^{+ =} present - = absent

Antimicrobial Efficacy of O. gratissimum Ethanolic Extract against Test Bacterial Isolates

The result of the *O. gratissimum* ethanolic leaf extract against *S. aureus, K. pneumoniae, P. aeruginosa, E. coli, S. marcescens* and *B. cereus* is shown in Table 2. All the isolates were susceptible to the extracts even at low concentrations without being resistant to any of the isolates. Ethanol extract produced an inhibition range of 15-35mm, 8-27mm, 7-26mm, 9-27mm, 5-24mm and 13-32mm at 12.5 to 200mg/ml *against S. aureus, K. pneumoniae, P. aeruginosa, E. coli, S. marcescens* and *B. cereus* respectively (**Table 2**).

Table 2: Antibacterial Activity of *O. gratissimum* Ethanolic Leaf Extract against Test Bacterial Isolates

	Zones	of inhib	oition (1	nm)	
Test isolates	200mg /ml	100mg /ml	50mg/ ml	25mg/ ml	12.5mg /ml
S. aureus	35±0.5	29±0.5	25±1. 0	19±1. 0	15±0.0
K. pneumonia e	27±1.1	21±1.0	18±0. 0	13±0. 0	8±0.4
P. aeruginos a	26±0.7	22±1.0	17±0. 5	10±0. 0	7±0.1
E. coli	27±0.0	22±0.0	18±0. 5	15±1. 1	9±0.0
S. marcescen s	24±0.0	20±0.4	14±1. 2	9±0.0	5±0.1
B. cereus	32±0.0	27±0.0	24±1. 2	18±0. 0	13

Antimicrobial Efficacy of *O. gratissimum* Methanolic Extract against Test Bacterial Isolates

Methanolic extract exerted inhibition in the range of 12-33mm, 11-31mm, 8-23mm, 12-25mm, 8-26mm and 6-26mm at 12.5 to 200mg/ml against S. aureus, K. pneumoniae, P. aeruginosa, E. coli, S. marcescens and B. cereus respectively (Table 3).

Table 3: Antibacterial Activity of *O. gratissimum* Methanolic Leaf Extract against Test Bacterial Isolates

Zone of inhibition					
Test isolates	200mg/ ml	100mg/ ml	50mg/ ml	25mg/ ml	12.5mg/ ml
S. aureus	33±1.1	30±1.0	25±0.0	20±0.0	12±0.5
K. pneumoniae	31±0.0	24±1.0	20±0.0	14±0.4	11±0.1
P. aeruginosa	23±1.2	19±1.0	16±0.0	12±0.5	8±0.0
E. coli	25±1.0	20±0.0	17±0.5	15±0.0	12±0.1
S. marcescens	26±1.0	22±1.0	17±0.0	13±1.0	8±0.0
B. cereus	26±0.0	23±0.5	14±1.0	10±0.0	6±0.0

Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) of *O. gratissimum* extracts

Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentrations (MBC) of O. gratissimum extracts on multi-antibiotic resistant S. aureus, K. pneumoniae, P. aeruginosa, E. coli, S. marcescens and B. cereus are presented in Table 4. The MIC of the ethanolic extracts were 12.5, 25, 25, 25, 12.5 and 50mg/ml on S. aureus, K. pneumoniae, P. aeruginosa, E. coli, S. marcescens and B. cereus respectively while methanol exerted MIC of 25, 50, 50, 12.5, 25 and 25mg/ml on S. aureus, K. pneumoniae, P. aeruginosa, E. coli, S. marcescens and B. cereus respectively. The MBC of the isolates on ethanolic extract was 100mg/ml for S. aureus, K. pneumoniae, P. aeruginosa and B. cereus, and 50mg/ml for E. coli and S. marcescens while the MBC on methanolic extract was 50mg/ml against E. coli, 100mg/ml against S. aureus, K. pneumoniae, P. aeruginosa, B. cereus and S. marcescens.

Table 4: Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) of Ethanolic and Methanolic *O. gratissimum* Leaf Extracts

	N	MBC (mg/ml)		
Test isolates	Ethanol	Methanol	Ethanol	Methanol
S. aureus	12.5	25	100	100
K. pneumoniae	25	50	100	100
P. aeruginosa	25	50	100	100
E. coli	25	12.5	50	50
S. marcescens	12.5	25	50	100
B. cereus	50	25	100	100

DISCUSSION:

The rate at which pathogens are becoming resistant to more antimicrobial agents and the diseases caused by multi-drug resistant bacteria cannot be exaggerated. Thereby, the need to explore nature for alternatives, either for direct usage or synthesized from nature. Plants are well known as the major reservoir of chemical agents used by pharmaceutical industries to produce drugs used for the treatment of various diseases.^[20] Also, several studies have been conducted on the therapeutic potential of various plant metabolites on disease-causing pathogens. The problem of antibiotic resistance is increasing and actions such as control measures in the use of antibiotics to treat diseases and improve livestock production should be promulgated as a high percentage of human disease-causing pathogens are gotten from animals.

In this study, we evaluated the phytochemical components of both methanol and ethanol extracts of scent leaf, the antibacterial activity of *O. gratissimum* extracts against multi-drug resistant bacteria and determined the least concentrations of both extracts required to kill and inhibit the bacterial growth. The phytochemical analysis revealed the absence of

anthraquinones in ethanolic extract, and terpenoids and phlobatannins in methanolic extract. These phytochemicals present in *O. gratissimum* leaves can be attributed to the plant's antibacterial activity. [21] Thereby, the reason for being used in the locality for treatment of many bacterial diseases. The finding on the presence of these bioactive agents in *O. gratissimum* ethanolic extract agrees with previous studies on the plant extract. [22, 23] These bioactive components have been reported by researchers to possess beneficial purposes to humans such as antitumor, anti-diuretic and treat gastrointestinal disorders. [24, 25]

The antibacterial analysis of the O. gratissimum extracts revealed that the extracts possess antibacterial efficacy against S. aureus, K. pneumoniae, P. aeruginosa, E. coli, S. marcescens and B. cereus by production of inhibition to their growth in in-vitro analysis. The extracts were found to have the highest inhibition on S. aureus. Despite the efficiency, the antibacterial activity of the extracts decreased with decrease in the concentration of the extracts which agrees with the previous studies on O. gratissimum. [22, ²³ O. gratissimum extracts have also been reported to possess antibacterial ability against E. coli, S. aureus and K. pneumonia, E. coli, Salmonella typhi and Yersinia enterolitica. [26, 27] The extracts showed varied potential against the test isolates with ethanol being more efficient on Gram-positive than Gram-negative bacteria. The susceptibility of these isolates to these extracts shows the reason for their use for the treatment of gastrointestinal infections, cough, and sore throat. Various literature and the results of this research work have shown the therapeutic potential of this plant in treating diseases. It is interesting to reveal that even at low concentration of 12.5mg/ml, O. gratissimum extracts were effective against B. cereus; a known spore-forming bacterium and recalcitrant to harsh environmental conditions. Also, the extracts showed bacteriostatic and bactericidal ability against S. aureus, K. pneumoniae, P. aeruginosa and E. coli that have become multi-drug resistant. This study showed Gram-positive bacteria to be more susceptible to the extracts than Gram-negative bacteria. Past studies also arrived at the same conclusion about Gram-negative bacteria being more resistant to plant extracts and antibiotics than Gram-positive bacteria. Plants exhibit antimicrobial activity on pathogens through mechanisms such as membrane disruption, inhibition of nucleic acids, proteins, and phospholipid synthesis. [29]

The MIC and MBC for both extracts ranged between 12.5-50mg/ml and 50-100mg/ml respectively. The possession of low MIC and MBC values by *O. gratissimum* extracts against these isolates was also revealed in a similar pattern by researchers. [21, 22, 26] These possession of high inhibition zones at low concentration, low MIC, and low MBC values by *O. gratissimum* extracts in this study against these pathogens show its medicinal ability in the treatment of diseases caused by the pathogens. This study therefore revealed the extracts of *O. gratissimum* to possess useful phytochemicals, low MIC, low MBC, and antibacterial ability on the selected infectious pathogens.

CONCLUSION:

The results of this study revealed that *O. gratissimum* extracts possess bioactive substances that are responsible for its antibacterial activity, showed high antibacterial activity against the multi-drug resistant bacteria used in this study and could be used to treat various infections caused by these pathogens in animal husbandry and humans.

RECOMMENDATION:

The extracts of *O. gratissimum* leaves are recommended for use in treating infections caused by *S. aureus, P. aeruginosa, E. coli, K. pneumoniae, B. cereus* and *S. marcescens*.

REFERENCES:

[1]. Yedjou C, Izevbigie E, Tchounwou P. Preclinical assessment of *Vernonia amygdalina* leaf extracts as DNA damaging anti-cancer agent in the

- management of breast cancer. *Inter. J. Environ. Res. Public Health* 2008; 12:123–127.
- [2]. Evbuomwan L, Chukwuka EP, Obazenu EI, Ilevbare L. Antibacterial activity of *Vernonia amygdalina* leaf extract against multi-drug resistant bacterial isolates. *J. Appl. Sci. Environ. Manage.* 2018;22(1):17–21.
- [3]. Anibijuwon II, Udeze AO. Antimicrobial activity of *Carica papaya* (pawpaw leaf) on some pathogenic organisms of clinical origin from South-Western Nigeria. *Ethanobotanical Leaflets* 2009; 13:850–864.
- [4]. Zhanar A, Aknur T, Ardak J, Kairolla R, Aigul J. Study of component composition and antimicrobial activity of the ophthalmic emulsion based on the safflower flowers (*Carthamus tinctorius* L.). *International Journal of Microbiology* 2022; 11:24–33.
- [5]. Akinjogunla OJ, Adegoke AA, Udokang IP, Adebayo-Tayo B. Antimicrobial potential of *Nymphaea lotus* (Nymphaeaceae) against wound pathogens. *Journal of Medicinal Plants Research* 2009;3(3):138–141.
- [6]. Abdullhai M. Phytochemical constituents and antimicrobial and grain protectant activities of clove basil (*Ocimum gratissimum* L.) grown in Niger. *Journal of Plant Research* 2012;2(1):51–58.
- [7]. Kpodekon MT, Boko KC, Mainil JG, Farougou S, Sessou P, Yehouenou B, Gbenou J, Duprez JN, Bardiau M. Composition chimique et test d'efficacité in vitro des huiles essentielles extraites de feuilles fraîches du basilic commun (Ocimum basilicum) et du basilic tropical (Ocimum gratissimum) sur Salmonella enterica sérotype Oakland et Salmonella enterica sérotype Legon. Journal de la Société Ouest Africaine de Chimie 2013; 35:41–49.
- [8]. Soro S, Abo K, Kone D, Coffi K, Kouadio JY, Ake S. Comparaison de l'efficacité antifongique de l'huile essentielle d'*Ocimum gratissimum* L. et du fongicide de synthèse mancozebe contre le mycopathogène tellurique, *Fusarium oxysporum* f. sp. *Radicis-lycopersici* en cultures de tomate

- (Lycopersicon esculentum Mill.) sous abri en Côte d'Ivoire. Agronomie Africaine 2011;23(1):43–52.
- [9]. Koane JN, Ouamba JM, Syssa-Magale JL. Études phytochimiques et pharmacologiques de quelques plantes médicinales centrafricaines à propriétés antidiabétiques. *Diabetes and Metabolism* 2012;38: A112–A113.
- [10]. Matasyoh LG, Josphat CM, Francis NW, Miriam GK, Anne WTM, Titus KM. Chemical composition and antimicrobial activity of the essential oil of *Ocimum gratissimum* L. growing in Eastern Kenya. *Afri. J. Biotechnol.* 2007; 6:760–765.
- [11]. Ogundare AO. Antibacterial properties of the leaf extract of *Vernonia amygdalina*, *Ocimum gratissimum*, *Corchorus olitorius* and *Manihot palmate*. *Journal of Microbiology and Antimicrobial* 2011;3(4):77–86.
- [12]. Rabelo M, Souza EP, Soares PMG, Miranda AV, Matos FJA, Criddle DN. Antinociceptive properties of the essential oil of *Ocimum gratissimum* L. (Labiatae) in mice. *Brazilian J. Med. Biol. Res.* 2003; 36:521–524.
- [13]. Prajapati ND, Purohit SS, Sharma AK, Kumar T. *Agro's Dictionary of Medicinal Plants*. 1st ed. Agrobios: India, 2003.
- [14]. Effraim KD, Jacks TW, Sopio OA. Histopathological studies on the toxicity of *Ocimum gratissimum* leaves extracts on some organs of rabbits. *African Journal of Biomedical Research* 2003; 6:21–25.
- [15]. Kouadio NJ, Guessennd NK, Kone MW, Moussa B, Koffi YM. Evaluation de l'activité des feuilles de *Mallotus oppositifolius* (Geisel.) Müll.-Arg (Euphorbiaceae) sur des bactéries multirésistantes et criblage phytochimique. *International Journal of Biological and Chemical Sciences* 2015;9(3):1252–1262.
- [16]. AMC. Analytical Methods Committee. Understanding and Acting on Scores Obtained in Proficiency Testing Schemes. Royal Society of Chemistry, 2002.

- [17]. Odeyemi AT, Fagbohun ED. Antimicrobial activities of the extracts of the peels of *Dioscorea* cayenesis L. J. Appl. Environ. Sci. 2005; 1:37–42.
- [18]. CLSI. Clinical Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility Testing. 30th ed. CLSI supplement M100, Wayne, PA, USA, 2020.
- [19]. Ajaiyeoba EO, Onocha PA, Noozo SO, Sama W. Antimicrobial and cytotoxicity evaluation of *Buchholzia coriacea* stem bark. *Journal of Agricultural and Environmental Science* 2003; 74:706–709.
- [20]. Nwachukwu M, Dike RN, Nwachukwu IO, Ohalete CN, Anyanwu R. Preliminary phytochemical assessment of extract from leaves and root of *Vernonia amygdalina* on *E. coli. World Journal of Pharmacy and Pharmaceutical Sciences* 2012;2(1):44–51.
- [21]. Umamaheswari A, Shree VR, Aparna N. In vitro antibacterial activity of *Bougainvillea* spectabilis leaves extracts. *Advance Biology* Research 2008; 2:1–5.
- [22]. Ojokoh A, Ojo MA. Antibacterial efficacy of *Ocimum gratissimum* on multidrug resistant *Staphylococcus aureus* and *Klebsiella pneumoniae* isolated from clinical and environmental samples in Ondo State. *Int. J. Med. Lab. Res.* 2018;3(3):39–51.
- [23]. Kin A, Abubakr I, Yaki LM, Olusola LF. Antibacterial efficacy of *Ocimum gratissimum* on some pathogenic gastrointestinal bacteria. *Africa Journal of Microbiology Research* 2018;12(40):923–929.
- [24]. Okwu DE. Phytochemicals and vitamin content of indigenous spices of South-eastern Nigeria. *Journal of Agricultural and Environmental Science* 2004;6(1):30–37.
- [25]. Khoobchandani M, Ojeswi BK, Ganesh N, Srivastava MM, Gabbanini S, Matera R, Iori R, Valgaimili L. Antimicrobial properties and analytical profile of traditional *Eruca sativa* seed oil: Comparison with various aerial and root plant extracts. *Journal of Antimicrobial Chemotherapy* 2010; 120:217–224.

- [26]. Ubafie MO, Ejale AU. Antimicrobial activity and quantitative analysis of *Ocimum gratissimum* on some pathogenic bacteria. *Issues in Biological Sciences and Pharmaceutical Research* 2008;6(2):17–22.
- [27]. Junaid SA, Olabode AO, Onwuliri FC, Okworiu AE, Agina SE. The antimicrobial properties of *Ocimum gratissimum* extracts on some selected gastrointestinal bacterial isolates. *Afr. J. Biotechnol.* 2006;3(22):2315–2321.
- [28]. Burt S. Essential oils: Their antibacterial properties and potential application in foods—a review. *International J. of Food Microbiology* 2004; 94:223–253.
- [29]. Walsh SE, Maillard JY, Russel AD, Catrenich CE, Charbonneau AL, Bartolo RG. Activity and mechanism of action of selected biocidal agents on Gram-positive and Gram-negative bacteria. *J. Appl. Microbiol.* 2003; 94:240–247.
- [30]. Okwu DE. Phytochemicals and vitamin content of indigenous spices of South-eastern Nigeria. *Journal of Agricultural and Environmental Science* 2004; 6(1):30–37.
- [31]. Khoobchandani M, Ojeswi BK, Ganesh N, Srivastava MM, Gabbanini S, Matera R, Iori R, Valgaimili L. Antimicrobial properties and analytical profile of traditional *Eruca sativa* seed oil: Comparison with various aerial and root plant extracts. *Journal of Antimicrobial Chemotherapy* 2010; 120:217–224.
- [32]. Ubafie MO, Ejale AU. Antimicrobial activity and quantitative analysis of *Ocimum gratissimum* on some pathogenic bacteria. *Issues in Biological Sciences and Pharmaceutical Research* 2008; 6(2):17–22.
- [33]. Junaid SA, Olabode AO, Onwuliri FC, Okworiu AE, Agina SE. The antimicrobial properties of *Ocimum gratissimum* extracts on some selected gastrointestinal bacterial isolates. *Afr. J. Biotechnol.* 2006; 3(22):2315–2321.
- [34]. Burt S. Essential oils: their antibacterial properties and potential application in

foods—a review. *International J. of Food Microbiology* 2004; 94:223–253.

[35]. Walsh SE, Maillard JY, Russel AD, Catrenich CE, Charbonneau AL, Bartolo RG. Activity and mechanism of action of selected biocidal agents on Gram-positive and negative bacteria. *J. Appl. Microbiol.* 2003; 94:240–247.

Cite of article: Adebesin IO, Kolapo KT, Amoka SO, Olubunmi OH, Olomoko CR, Balogun AO, Ganiyu LT. Phytochemical analysis and antibacterial efficacy of *Ocimum gratissimum* extracts on multi-drug-resistant bacteria. *Int. J. Med. Lab. Res.* 2023;8(3):23–31. http://doi.org/10.35503/IJMLR.2023.8304

CONFLICT OF INTEREST: Authors declared no conflict of interest

SOURCE OF FINANCIAL SUPPORT: Nil

International Journal of Medical Laboratory Research (IJMLR) - Open Access Policy

Authors/Contributors are responsible for originality of contents, true references, and ethical issues.

IJMLR publishes all articles under Creative Commons Attribution- Non-Commercial 4.0 International License (CC BY-NC). https://creativecommons.org/licenses/by-nc/4.0/legalcod